1. Evolutionary History of the Nitrogen, Carbon, and Oxygen Cycles

1.1. Nitrogen in the preoxygenated world/oceans

All organisms on Earth are composed primarily of the six light elements: H, C, N, O, S, and P. Hydrogen was created 13.7 Ga (billion years ago) in the “Big Bang” while the other five elements were created via fusion reactions in stars. Their relative...
abundance scales, to a first order, with the inverse of their atomic mass. Hence, among these five elements in our solar system, H is the most abundant and P is the rarest (Williams and Frausto Da Silva, 1996). When our solar system was formed, ca. 4.6 Ga, a complement of elements was imported from supernovae and distributed among the planets and planetary bodies (e.g., moons and asteroids) and our sun. During planetary accretion, formation of dense phases caused siderophile elements (e.g., Fe metal) to move towards the core, while lighter elements tended to migrate toward the mantle or the atmosphere.

In early models of planetary accretion, the process was thought to be relatively slow, such that elements such as N and C had high probabilities of combining with H to form highly reduced molecules such as NH₃ and CH₄ (Urey, 1952). However, based on more detailed observations of how planets are formed, as well as on theoretical calculations about the rate of outgassing of H₂ to space, a much larger fraction of C and N probably existed in higher oxidized states such as CO₂ and N₂ (Holland, 1984), but almost certainly not NOₓ. Chemically, N₂ is highly inert. The biologically accessible form of the element requires that it be used in dissimilatory metabolism (such as bacterial conversion from nitrate to nitrite) or react either at a lower reducing level, with H, to form NH₃, or at a higher oxidized level, NOₓ, from which it can be biologically reduced to NH₃. Assuming the described initial conditions, the environment of the Archean oceans exerted a strong selection pressure for processes that could reduce N₂ to NH₃, thereby making N₂ a sink for electrons in an electron market where oxidized species are relatively rare. Under such conditions, biological reduction of N₂ could serve as a pathway for the oxidation of organic carbon under anaerobic conditions, or as a detoxification for cyanides, which could have been common in the ancient oceans (Fani et al., 2000; Postgate, 1998). This redox couple, namely the oxidation of organic carbon and the reduction of N₂, is sensitive to oxygen, and hence the biologically-mediated geochemical cycles of the three elements, C, N, and O, are inextricably interconnected (Box 35.1).

In this chapter we examine the connections between these three elements and focus on biological nitrogen-fixation to illustrate the regulatory control of biological processes on the geochemistry of the oceans and Earth over the past 4 billion years.

1.2. Oxygenation of the early atmosphere: The influence of shelf area and nitrogen fixation rates

Geochemical evidence suggests that there were delays of several hundred million years between the rise of oxygenic photosynthesis, the oxygenation of the atmosphere, and the oxygenation of the deep ocean. Photosynthesis (evidenced by cyanobacterial microfossils and biomarkers) rose as early as 3.5 Ga (billion years before present; (Schopf, 1993)) and had been solidly established by 2.7–2.5 Ga (Brocks et al., 1999; Knoll, 1996; Schopf, 1993; Summons et al., 1999). Data from red beds, detrital mineral deposits, and sulfur isotopes indicate the rise of atmospheric oxygen around 2.4 Ga (Bekker et al., 2004; Chandler, 1980; Des Marais et al., 1992;
The deep ocean appears to have remained anoxic at least until the disappearance of banded iron formations at 1.8 Ga (Holland, 1984), and possibly as late as 0.8 Ga (Arnold et al., 2004; Canfield, 1998; Canfield and Teske, 1996).

It is likely that oxygen feedbacks on N and C cycles substantially delayed the rise of atmospheric oxygen. This hypothesis is supported by box model calculations, incorporating ocean circulation and ocean-atmosphere gas-exchange with the geochemical cycles of C, N, and O (Fennel et al., 2005; Fig. 35.1). The model has four ocean boxes (high and low latitude surface, deep ocean and shelf seas) coupled to a single atmospheric reservoir and represents the nitrogen and oxygen cycle interactions excluding the sedimentary processes (Fig. 35.1). Application of this model shows that, when initialized with anaerobic conditions, ammonium is originally stable in the reduced

Box 35.1 Links Between Oceanic Nitrogen and Oxygen Cycles

The oceanic cycles of nitrogen and oxygen are intimately linked through the processes of nitrification and denitrification. These processes constitute the key source and sink of fixed nitrogen in the ocean and control its global budget. The coupling of the nitrogen and oxygen cycles has implications for the long-term chemical evolution of the ocean and the atmosphere, variations in the oceans’ nitrogen budget, and the nitrogen limitation of marine ecosystems over centennial to very long timescales.

During photosynthesis nutrients, including nitrogen, are assimilated into organic matter and molecular oxygen is produced, depicted schematically by

\[
106\text{CO}_2 + 16\text{NO}_3^- + \text{H}_2\text{PO}_4^- + 122\text{H}_2\text{O} \rightarrow \text{C}_{106}\text{H}_{263}\text{O}_{110}\text{N}_{16}\text{P} + 138\text{O}_2 \quad (35.1)
\]

Here we assume a source of fixed nitrogen in the form of nitrate. Alternatively, diazotrophs can fix their own nitrogen, reducing N\textsubscript{2} gas to the level of ammonium in order to be assimilated for protein synthesis. Ammonium may be oxidized by nitrifying bacteria to produce nitrite and nitrate and consuming molecular oxygen:

\[
\text{NH}_4^+ + 2\text{O}_2 \rightarrow \text{NO}_2^- + 2\text{H}_2\text{O} \quad (35.2)
\]

The sequence of nitrogen fixation and nitrification is a source of fixed nitrogen and a sink of dissolved oxygen in the ocean. Aerobic respiration of organic material by heterotrophic bacteria also consumes oxygen and releases nitrogen to the dissolved inorganic pool; the reverse of (35.1).

Under very low oxygen conditions, denitrification, resulting from the anaerobic reduction of nitrate by heterotrophic bacteria, consumes fixed nitrogen in the form of nitrate, ultimately regenerating N\textsubscript{2} gas (Froelich et al., 1979).

\[
\text{C}_{106}\text{H}_{263}\text{O}_{110}\text{N}_{16}\text{P} + 84.8\text{HNO}_3^- \rightarrow 106\text{CO}_2 + 42.4\text{N}_2 + 16\text{NH}_3 + \text{H}_3\text{PO}_4^- + 148.4\text{H}_2\text{O} \quad (35.3)
\]

Hence, the sources and sinks of dissolved inorganic nitrogen and dissolved oxygen in the ocean are intimately linked and their global cycles coupled.
deep ocean. Yet, as free oxygen becomes available in the ocean, ammonium is converted to nitrate, which can be rapidly reduced to N₂, thus decreasing the pool of fixed inorganic nitrogen. Hence, the interaction between N, C, and O during a transition from an anaerobic to aerobic ocean leads to a strong negative feedback. During this low-nitrogen stage, which can last several 100 million years in our simulations, export production is severely attenuated. Unless some exported carbon is buried, oxygen cannot accumulate in the upper ocean or atmosphere.

Figure 35.1 Schematic depiction of the coupled, global ocean nitrogen and oxygen cycles. Dissolved inorganic nitrogen and nitrogen fixation fuel oxygenic photosynthesis in the surface ocean, helping to sustain atmospheric and deep ocean oxygen levels. In the deep ocean, oxygen is typically consumed during the microbial respiration of organic matter, producing dissolved inorganic nitrogen. However, in low oxygen waters and sediments, denitrification fuels the oxidation of organic matter, ultimately removing dissolved inorganic nitrogen from the water column. In the early ocean, initial oxygenation of the water column may have inhibited the build up of nitrate which would have been quickly consumed as the oxidant of organic matter.
The net rate of oxygen production during the low (fixed)-nitrogen stage at the anoxic/oxic transition depends on several factors, including the burial of organic matter and the rate of oxygen consumption during nitrification. In our model studies, the oxygenation of the ocean and atmosphere critically depend on the presence of shallow continental shelf seas (Fennel et al., 2005). The burial of organic matter is much more efficient in shallow continental shelves than in the deep ocean (Hartnett et al., 1998) hence, rates of organic matter burial and net oxygen production should increase as a function of the relative area of continental shelves and shallow seas (Berner, 1984). The relationship between the area of continental shelves, carbon burial, and oxygen production highlights the fundamental role of tectonic processes, especially seafloor spreading, in controlling eustatic sea level and the area of shallow seas (Miller et al., 2005), and in determining the rate of oxygen evolution. The model also demonstrates that an increase in the oceanic phosphate inventory will enhance the net rate of oxygen production owing to higher export. In a fully oxidized ocean, it also potentially leads to an increased inventory of fixed inorganic nitrogen. Furthermore, the rate of net oxygen production is very sensitive to the N2 fixation rate, which suggests that the initial increase of oxygen had an additional negative feedback on oxygen production by compromising nitrogenase activity both directly and through trace metal limitation (Anbar and Knoll, 2002; Falkowski, 1997) (see below and Fig. 35.2).

An extended period of severe nitrogen limitation and low productivity during the Proterozoic is consistent with the δ13C record of carbonates, which indicates large variations at the beginning and end of the Proterozoic, but not in the intervening period (Des Marais et al., 1992). Our model studies suggest that during the enhanced tectonic activity at the end of the Proterozoic, changes in the global shelf area or in the oceanic phosphate inventory could have triggered the oxygenation of the deep ocean.


2.1. Evolutionary constraints for nitrogen fixation and adaptive strategies

Over a 200 million year period, centered around 2.3 billion years ago (Bekker et al., 2004), the partial pressure of oxygen (pO2) rose from <4 × 10^-6 atm to >0.03 atm (Pavlov and Kasting, 2002; Rye and Holland, 1998). This emerging environment especially challenged diazotrophic cyanobacteria, the only diazotrophs capable of oxygenic photosynthesis, yet having to contend with the innate sensitivity of the nitrogenase enzyme to molecular oxygen. Diazotrophic cyanobacteria provide an elegant case-study of how the interplay between the N, C and O cycles influenced the adaptation of key metabolic processes, and how these cellular processes feed back
Figure 35.2  (A) Northern blots of nif HDK transcript abundance for *Trichodesmium* IMS101 cultures bubbled with nitrogen (0% oxygen), 5%, 20%, and 50% O<sub>2</sub> for 5 h. (B) Western blots of nitrogenase protein abundance (challenged with universal Fe-protein polyclonal antibodies) for the above experiment (C) Nitrogenase activity (presented as % of air-control) measured by acetylene reduction for the above experiment 1 h after induction of bubbling. (D) Relationship between O<sub>2</sub> concentrations and nitrogen fixation (% from maximum rates) for *Trichodesmium* IMS101 and other diazotrophs. Filled triangles: experimental data from our cultures of *Trichodesmium* IMS101; empty triangles with cross—*Trichodesmium* NIBB1067 (Ohki and Fujita, 1988), empty triangles—*Trichodesmium* spp. field populations (Mague et al., 1977; Saino and Hattori, 1982), black circles—*Anabaena cylindrica* (Gallon et al., 1993), black squares—*Gloeothecae* (Nageli) (Gallon et al., 1993). Short-term (1–2 h) anaerobic incubation yields maximum nitrogenase activity for aerobic diazotrophs (C). On longer time scales, respiratory requirements will yield maximum activity (*in vivo*) at microaerobic oxygen concentrations (D).
into the aquatic and global systems. In this section we focus on the evolutionary constraints on nitrogen-fixation in cyanobacteria. We then discuss how these biological controls feed back into the geochemical cycles.

The evolutionary roots of nitrogen fixation, inferred from molecular phylogenetic analyses of $\textit{nifH}$ genes, suggest that the genes encoding for nitrogenases are ancient and underwent horizontal gene transfer, gene duplication, recruitment and fusion (Raymond et al., 2004). Indeed, phylogenetic analyses do not rule out the possibility that nitrogenase was present in the last universal common ancestor (LUCA) and, therefore predates the divergence of the archaea and bacteria (Fani et al., 2000; Raymond et al., 2004; Woese, 1998), or if it evolved in methanogenic archaea and was subsequently transferred to a primitive bacterium (Raymond et al., 2004). In either scenario, the catalytic subunits of the enzyme complex clearly indicate that the original enzyme existed long before the oxygenation of Earth’s atmosphere (Broda and Pesheck, 1983) and that trace elements such as Fe, Mo, and V additionally modulated its evolutionary trajectory.

The influence of required trace elements on the evolution of nitrogenase has been discussed by several authors (Anbar and Knoll, 2002; Kustka et al., 2003; Raven, 1988; Raymond et al., 2004). The availability of iron influences $\text{N}_2$ fixation in diazotrophic cyanobacteria, from its direct effect on Fe-rich protein synthesis such as nitrogenase, to effects on photosynthesis, growth, and global productivity (Berman-Frank et al., 2001, 2007; Falkowski, 1997; Kustka et al., 2003; Mills et al., 2004; Paerl et al., 1994; Reuter et al., 1988). In the anaerobic environments of the Archean oceans, Fe would have been found predominantly in its reduced form (FeII) rather than FeIII (Anbar and Knoll, 2002 and Table 1 in Berman-Frank et al., 2003). The necessity for nitrogen selected for nitrogenase, while the availability of reduced iron selected for the metal complement of nitrogenase. Both laboratory studies (Berman-Frank et al., 2001; Berman-Frank et al., 2007, Shi et al., 2007; Kupper et al., 2008) and field simulations (Mills et al., 2004) show a direct link between the availability of nitrogen fixation rates in environmentally important diazotrophs (see Section 3.1). Thus, diazotrophy and subsequent primary production were predicted and found to be limited in the iron deficient regions of the subtropical and tropical oceans, such as the contemporary South Pacific (Moutin et al., 2008; Raimbault and Garcia, 2008).

Molybdenum (Mo) is another essential metal co-factor of nitrogenase. Mo, a rare trace element is much more abundant in the contemporary ocean than under the anaerobic conditions prevailing when nitrogenase first evolved, and is also required in the bacterial reduction of nitrate to nitrite (Williams and Frausto Da Silva, 2002). In the anoxic, asulfidic Archean oceans, up to 90% of the Mo would have been complexed in relatively insoluble sulfide minerals in igneous rocks (Anbar and Knoll, 2002; Falkowski, 2001; Williams and Frausto Da Silva, 1996). Limited availability of Mo may have been exacerbated in the mid-Proterozoic, where weathering, under a moderately oxidizing atmosphere, would have enhanced the delivery of sulfate ($\text{SO}_4^{2-}$) and molybdate to the deep ocean. Combined with primary production in the surface waters, this would have resulted in extremely high $\text{H}_2\text{~S}$ concentrations and removal of Mo via increased precipitation and formation of active thiomolybdate ($\text{MoS}_4^{2-}$) (Anbar and Knoll, 2002; Arnold et al., 2004).
Whereas alternative, older nitrogenases exist, in which Fe or V were replaced by Mo, these forms are catalytically less efficient. Interestingly, Mo-independent nitrogenases have been found only in heterocystous diazotrophs (phylogenetically the most recently diverging group—see discussion below) but not in non-heterocystous species (Bergman et al., 1997).

Past depletions in dissolved inorganic phosphate may have also limited nitrogen-fixation and primary production due to high adsorption of phosphate by iron-rich minerals. Indeed, based on adsorption isotherms in banded iron formations, sea-water phosphate concentrations in the Archean have been suggested to have been more than ten fold lower than the present values (0.15–0.6 µM versus the modern value of 2.3 µM) (Bjerrum and Canfield, 2002). Thus, present restrictions of phosphate on N₂ fixers in some oligotrophic areas such as the western subtropical Atlantic (Mills et al., 2004) and in the tropical Pacific (Moutin et al., 2007) may have correlations in the paleoceanographic records of the late Archean eon.

Nitrogen fixation is metabolically costly, requiring relatively large inputs of energy, reducing power, ancillary antioxidant enzymes and metal co-factors such as iron and Mo (Raven, 1988; Kustka et al., 2003). A priori, the nitrogenase enzyme is notorious for its sensitivity to molecular oxygen and for the irreversible damage to the the Fe₄S₄ clusters, in-vitro (Burgess and Lowe, 1996). Moreover, high oxygen stress causes proteolysis of nitrogenase subunits (Durner et al., 1996), suppresses nitrogenase synthesis, and leads to a shortage of respiratory substrates and reductants necessary for nitrogen fixation and assimilation (Gallon, 1992). Inhibitory effects of moderate oxygen levels, or short exposure times, in-vivo may be reversed, leading to an increase in nitrogen fixation rates (Gallon, 1992; Ludden and Roberts, 1995; Pan and Vessey, 2001; Prosperi, 1994; Yakunin et al., 2001) and, in some diazotrophs, to post-translational modification of the Fe protein from an inactive to active form (Ernst et al., 1990; Jouanneau et al., 1983; Ohki et al., 1991; Sweet and Burris, 1982; Zehr et al., 1993).

The fundamental constraint of the two conflicting processes has led to several adaptations including spatial (e.g. heterocystous cyanobacteria) and temporal separation of photosynthesis and N₂ fixation (e.g. unicellular species such as Cyanothecce), or a combination of both (e.g. in the non-heterocystous colonial forms such as Trichodesmium). The oxygen protection mechanisms have been the focus of several reviews examining molecular, phylogenetic, physiological, morphological, regulatory, and evolutionary adaptations in cyanobacteria (Adams, 2000; Bergman et al., 1997; Berman-Frank et al., 2003; Bohme, 1998; Fay, 1992; Gallon, 1992, 2001; Tuli et al., 1996). The various strategies to overcome the inherent conflict between oxygenic photosynthesis and nitrogen fixation reflect the wide-ranging flexibility and niches occupied by cyanobacteria: from anaerobic sediments to pelagic waters saturated with oxygen. Molecular phylogenies, of N₂ fixers using the nifH gene and small subunit ribosomal RNA sequences, showed no correlation between the phylogenetic relationships and the type of N₂ fixation (Raymond et al., 2004; Turner et al., 2001). This lack of correlation suggests multiple gains and/or losses of N₂ fixation among the different cyanobacteria (Raymond et al., 2004; Turner et al., 2001). The loss of cyanobacterial nif genes implies that different strategies arose early in the evolution of the clade, where some organisms
were able to adapt to an oxic world, while others were not (e.g., Oscillatoria). Commonly, many phylogenies show the most recent phylogenetic branching (both for nifH and RNA trees) in filamentous species where complete segregation of N₂ fixation and photosynthesis was achieved with the evolution of heterocystous cyanobacteria (Wolk et al., 1994).

Heterocystous cyanobacteria are predominantly terrestrial, fresh water and coastal species, inhabiting eutrophic or brackish environments (e.g. the Baltic Sea), with a few epiphytic and symbiotic representatives in the marine environment (Paerl and Zehr, 2000) (see Chapter 4, Carpenter and Capone, this volume). Very few heterocystous free-living species are found in the pelagic oceans, though a novel species was found, designated Anabaena gerdii (Carpenter and Janson, 2001). The heterocyst glycolipid envelope may confer an advantage for cyanobacteria inhabiting temperate brackish and fresh waters. These waters are characterized by an increase in O₂ flux by ~25% from that in sea water, and thus the thick glycolipid layer can help decrease diffusion of oxygen into the heterocysts (Staal et al., 2003a). At higher temperatures, typical of tropical waters, the glycolipid envelope of the heterocyst does not provide additional protection against oxygen, which may explain the paucity of heterocystous free-living bacteria in such environments (Staal et al., 2003a).

In addition to specialized niche selection and avoidance strategies, whereby nitrogen fixation is spatially or temporally separated from photosynthesis, oxygen concentrations influence transcription, translation, and changes in nitrogenase activity of cyanobacteria. We demonstrate this with results from the bloom-forming cyanobacterium Trichodesmium IMS101, which contributes significantly to nitrogen fixation in the tropical and sub-tropical oceans. Sequence and structural analyses of its nitrogenase are similar to those of other diazotrophic organisms (Zehr et al., 1997), and there is a post-translational modification between the active and non-active form of the protein under natural and induced conditions (Ben-Porath et al., 1993; Ohki et al., 1991). The abundance of nifH transcripts and its corresponding gene product were assayed in Trichodesmium cultures incubated for 5 h with 0, 5, 21 (present atmospheric level—PAL), and 50% O₂ (Fig. 35.2A). The results indicate that extracellular O₂ does not significantly influence transcription or translation of nifH on this time scale (Fig. 35.2A and B). In contrast, nitrogenase activity was strongly depressed by O₂; at 50% O₂ >90% of the nitrogenase activity was inhibited within 1 h (Fig. 35.2C).

Post-translational modification of activity appears relatively common in diazotrophic cyanobacteria and operates at much lower concentrations of oxygen than those required for repression of nif genes (Dominic et al., 1998; Tuli et al., 1996). A compilation of published data on the response of nitrogen fixation to varying ambient O₂ concentrations suggests a general relationship that is consistent with both the short-term response of the enzyme to O₂ and the organisms’ longer evolutionary adaptations where O₂ is used as sink for electrons in respiration (Fig. 35.2D). On average, 20–30% of the enzymatic activity is inhibited at present atmospheric oxygen concentrations of 21% (Fig. 35.2D).

In Trichodesmium spp., maximum nitrogen fixation occurs at oxygen concentrations of 2.5–5% PAL. Under truly anaerobic conditions, the Embden–Myerhof
(glycolysis) pathway apparently cannot supply sufficient substrates through fermentative reactions to meet the demands of nitrogen fixation. Hence, in vivo, the activity of the enzyme declines. As oxygen concentrations increase above 5%, the enzyme activity is inhibited, with only ~70–80% activity measured at atmospheric oxygen concentrations of 21%. At oxygen concentrations greater than 30%, a sharp drop in activity reduces fixation efficiency (potential) to <10% within minutes. This pattern of an inefficient nitrogenase is corroborated in data from other aerobic cyanobacteria such as in the heterocystous *Anabaena* and in the unicellular *Gloeothecae* (Fig. 35.2D). In these cyanobacteria, lowering the oxygen below atmospheric concentrations also enhances relative nitrogenase activity at short durations. However, at very low oxygen concentrations, lack of respiratory production of energy and substrates ultimately affects nitrogen fixation. Thus, in *Trichodesmium* for example, low oxygen stress caused faster mortality and biomass crashes after ~48 h (Berman-Frank, unpublished).

### 2.2. Adaptive strategies. Oxygen consumption—The Mehler reaction

In cyanobacteria, oxygen is potentially consumed through aerobic respiration and two light-dependent reactions, the oxygenase activity of RuBisCO (photorespiration) and the photosynthetic reduction of O₂, termed pseudocyclic photophosphorylation or the Mehler reaction (Box 35.2). In contrast to terrestrial C3 plants, which have relatively high rates of photorespiration, photorespiration of oceanic phytoplankton is usually low when dissolved inorganic carbon concentrations in seawater are at equilibrium (~2 mM) with the atmosphere. Moreover, cyanobacteria operate a CO₂ concentrating mechanism (CCM) which raises the CO₂ concentration in the vicinity of RuBisCO and inhibits oxygenase activity (Kaplan and Reinhold, 1999).

The Mehler reaction is a photochemical reduction of O₂ to H₂O₂ or H₂O in photosystem I (Box 35.2). Mehler activity is thought to be a mechanism for energy dissipation under high light intensities or when carbon fixation is limited by supply

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**Box 35.2 The Mehler reaction (pseudocyclic photophosphorylation)**

In 1951, the late Alan Mehler (1951) observed that chloroplasts can use oxygen as an electron acceptor. The reaction sequence is

\[
\text{H}_2\text{O} + 2\text{O}_2 \rightarrow 2\text{O}_2 + 2\text{H}^+ + \frac{1}{2}\text{O}_2
\]

\[
2\text{O}_2 + 2\text{H}^+ \rightarrow \text{H}_2\text{O}_2 + \text{O}_2
\]

and, in the presence of catalase:

\[
\text{H}_2\text{O}_2 \rightarrow \text{H}_2\text{O} + \frac{1}{2}\text{O}_2
\]

Net gas exchange is absent since the overall electron transport reaction which involves both Photosystems II and I is:

\[
\text{H}_2\text{O} + 2\text{O}_2 \rightarrow \text{H}_2\text{O} + 2\text{O}_2
\]
of inorganic carbon (Helman et al., 2003). Since the products of O₂ reduction are often superoxide and hydrogen peroxide (with superoxide dismutase catalyzing the reduction of superoxide to peroxide), Mehler activity has been hypothesized to be a metabolic defect rather than an adaptive strategy (Patterson and Myers, 1973). However, in the cyanobacterium *Synechocystis* sp. (PCC 6803), superoxide is reduced directly to water without a hydrogen peroxide intermediate (Helman et al., 2003). This single step reduction of superoxide to water is catalyzed by A-type flavoproteins; two of which (flv1, flv3) were identified as essential for this activity (Helman et al., 2003). Examination of the genome of *Trichodesmium*, identifies homologous genes to flv1 and flv3 with 62% and 67% sequence identity, respectively (Milligan et al., 2007).

Mehler activity is generally considered a process which can only consume photosynthetically derived O₂, and it cannot cause net consumption of O₂ because PSI activity relies on photosynthetically derived electrons (Kana, 1993). Yet, the shared-arrangement of photosynthetic and respiratory electron transport chains in cyanobacteria allows electrons from respiratory derived NAD(P)H to feed into the plastoquinone pool of the photosynthetic electron transport chain and reduce PSI (Schmetterer, 1994). Through the translocation of reductant (i.e. glucose 6-phosphate) from cells with functional PSII, Mehler activity can result in a net consumption of O₂ in cells (or heterocysts) which have no PSII activity and in which nitrogen is fixed (Fig. 35.3).

Results from *Trichodesmium* provide an example of the Mehler pathway’s role in modulating oxygen and facilitating nitrogen-fixation. In this species, under nitrogen-fixation conditions, approximately 50% of gross O₂ production is consumed through Mehler activity (Fig. 35.4); this is about twice the rate reported for *Synechococcus* (~25% of gross O₂ production) exposed to photoinhibitory irradiances (Kana, 1992). Mehler activity is dependent both on the time of day and the nitrogen source (Fig. 35.4). The period of maximum N₂ fixation is coincident with a decline in the net production of O₂ and a rise in the consumption of oxygen via Mehler activity, which is consistent with the hypothesized role of this pathway as a mechanism to protect nitrogenase from O₂ damage. In nitrate-grown *Trichodesmium*

![Figure 35.3](image-url) Photosynthetic and respiratory electron transport chains in cyanobacteria showing the shared electron carriers of each pathway (based on Schmetterer, 1994).
cultures (with negligible nitrogen-fixation), Mehler activity increases with light induction, but quickly drops to low and constant rates (10% of gross production) through the rest of the photoperiod (Fig. 35.4).

In *Trichodesmium*, Mehler activity is essential, as *Trichodesmium* relies on short term regulation of PSII and nitrogenase activities to separate these functions within a trichome (Berman-Frank *et al.*, 2003). PSII activity is regulated on time scales of 10–15 min and appears to involve the association/disassociation of the light harvesting complex (LHC II) from PSII (Küpper *et al.*, 2004). Nitrogenase activity is also regulated on similar time scales when incubated at different oxygen concentrations, with transcriptional and translational regulation requiring longer time-scales and higher concentrations of exposure (Fig. 35.2). While PSII activity is repressed in N₂ fixing cells, the activity of PSI is responsible for net O₂ consumption relying on the translocation of reductant for the donor side of PSI and the flux of photons driving the oxidation. Cultures of *Trichodesmium* grown under low (5%) oxygen showed some nitrogenase activity during the night; this

Figure 35.4 Time course of oxygen production (□), consumption (▲) and acetylene reduction (○) during the photoperiod in *Trichodesmium* sp. IMS l01 grown with N₂ (A) and nitrate (B) as nitrogen sources. Error bars are ±1 standard deviation.
activity was absent in controls (21%) and in high (50%) oxygen cultures (Küpper et al., 2004). The lack of Mehler activity at night in the controls (21%) and in the high oxygen cultures may thus reduce the total possible oxygen consumption and prevent nitrogenase activity. At lower O$_2$ concentrations, N$_2$ fixation can proceed in darkness because the respiratory rates are sufficient to consume O$_2$.

### 2.3. Biological feedbacks to the global nitrogen, carbon, and oxygen cycles

Oxygen-consuming pathways, including the Mehler reaction, deliver a positive feedback to nitrogenase and nitrogen fixation by reducing the concentrations of molecular and reactive oxygen that inhibit the enzyme. Yet the presence of oxygen is essential as respiratory pathways provide the required substrates for subsequent nitrogen assimilation. Thus, oxygen concentrations will exert both a positive and negative feedback towards adaptations at the organism-scale, which reflect a compromise of opposing metabolic requirements. At the molecular/biochemical scale, oxygen concentrations impose an upper limit on the nitrogenase enzyme for all types of cyanobacterial diazotrophs. Whether the regulation is at the level of transcription, translation, or metabolic activity, depends on the concentration and duration of inhibition by oxygen. Chronic inhibition of nitrogenase at PAL of O$_2$ imposes a metabolic inefficiency that reflects an extraordinarily slow tempo of evolution for this critical biogeochemical process. The high conservation within the primary sequence of nif genes suggests that the evolutionary risks associated with modifying nitrogenase outweigh the costs of its production. Nitrogenase is analogous to other core metabolic proteins, such as the reaction center protein of photosystem II, D1 and ribulose 1,5 bisphosphate carboxylase/oxygenase (Rubisco), which also operate at a fraction of their capacities under ambient atmospheric conditions or undergo extremely high rates of turnover as a result of post-translational damage. The inhibition of all of these proteins, either directly by elevated O$_2$, or indirectly through reactive oxygen species, potentially exerts a strong biological control on the upper bound of the concentration of the gas in Earth’s atmosphere, leading to metabolic and biogeochemical inefficiency in N$_2$ fixation. The inefficiency imposes a major elemental taxation on diazotrophic cyanobacteria, both in the costs of protein synthesis and in the scarce trace elements, such as iron, which has, in turn, led to a global limitation of fixed nitrogen in the contemporary oceans (Falkowski, 1997).

We extended and applied the systems-analysis based approach of Beerling and Berner (Beerling and Berner, 2005) to the feedbacks between (aquatic) nitrogen fixation and the cycles of carbon and oxygen (Fig. 35.5). While the role of sustained forest fires in maintaining PAL of oxygen (Lenton and Watson, 2000; Watson et al., 1978) is currently debated (Wildman et al., 2004) (Fig. 35.5), nitrogen fixation (via the evolutionary sensitivity of nitrogenase) acts as an additional control, limiting the upper concentration of oxygen in the atmosphere. The reduction of nitrogen-fixation due to high oxygen concentrations could lead to a decline in both diazotrophic and total aquatic primary productivity (with parallel declines in the terrestrial biomass due to higher photorespiration under high oxygen), and to reduced rates of organic burial (Fig. 35.5 path 1–2–3). Lower burial rates would lead to a decrease in oxygen and an
3. Sensitivity of the Nitrogen, Carbon, and Oxygen Cycles to Climate Change

Biological feedbacks into the global nitrogen and oxygen cycles are intricately dependant on climate patterns and oceanic circulation which also influence trace metal availability in the upper ocean (Fig. 35.5). In this section we explore future scenarios in the oceanic nitrogen, carbon and oxygen cycles due to shifts in aeolian iron availability, variations in climate, and ocean circulation.

Figure 35.5 Simplified systems analysis diagram of the feedbacks between nitrogen fixation (mainly by aquatic cyanobacterial diazotrophs) and the oxygen and carbon cycles. Plain arrows show direct responses while arrows with bull’s-eyes illustrate inverse effects. The +/- indicates feedbacks could go in either direction depending on conditions. The feedback loop is designated by numbers adjacent to path. Note that the processes and feedbacks operate at different time-scales (from hours (e.g. N-fixation and photosynthesis) – to millenia (e.g. global O₂, CO₂ concentrations). Diagram modified from Beerling and Berner (2005).

increase in atmospheric CO₂ (Fig. 35.5 paths 1-2-3-4, 1-2-3-5). If oxygen concentrations are maintained at PAL or slightly lower (see Fig. 35.2D), nitrogen fixation may be sufficient to drive high total primary production and higher burial rates, and a negative feedback to both C and O concentrations (Fig. 35.5).
3.1. Aeolian iron and stratification of the surface mixed layer in the future

The major source of Fe in the surface waters of several large ocean regions is the aeolian dust transported from the continents (Duce and Tindale, 1991; Fung et al., 2000; Jickells et al., 2005). Yet the evidence for the relative importance of aeolian sources versus *in situ* iron supply by upwelling waters from depths is unclear (Coale et al., 1996). The distribution of aeolian Fe varies strongly with the seasons and from one ocean region to another. The predominant fraction of the Fe inputs enters the oceans in the Northern Hemisphere, where the high Fe fluxes are primarily concentrated in low and mid-latitudes (Gao et al., 2001). These are characterized by disproportionate seasonal flux rates, with summer Fe fluxes approximately twice those of winter fluxes (Gao et al., 2001). This non-uniform distribution feature is largely reflected by the distribution of dust sources (Ginoux et al., 2001; Zender et al., 2003). The current estimate of the total Fe deposition to the ocean is $14 \times 10^{12}$ g year$^{-1}$, which includes substantial uncertainties. The major processes that control the delivery of aeolian Fe to the ocean are dry deposition by gravitational settling of particles, turbulence in the surface layer of the atmosphere, and wet deposition through precipitation scavenging. Any variation in the strength and distribution of dust sources, and removal processes caused by climate change and land use, could affect dust emissions (Tegen et al., 2004) and consequent delivery of aeolian iron to the ocean (Fig. 35.5 path 8-13).

One example is the increasing concentrations of atmospheric CO$_2$ and enhanced global temperatures which have altered the global hydrological cycle. This is reflected by changes in precipitation patterns that may have profound impacts on dust emissions and subsequent deposition to the oceans. Over the past century there has been a global increase in rainfall over land (Dai et al., 1997). Enhanced precipitation over desert regions, in particular, has been considered as a negative feedback to desertification (Miller et al., 2004). The increased rainfall could directly impact the emissions of dust through altering soil moisture (aridity) in the source regions and dust burden in the atmosphere and, consequently, affect the concentrations of dust over the oceans. Observations made at Barbados in the tropical North Atlantic indicate that variations in dust concentrations from 1965 to 1998 were inversely correlated with rainfall in the Soudano-Sahel region in Africa (Prospero and Lamb, 2003). This finding suggests that atmospheric dust loading over the ocean is sensitive to the strength of precipitation in the dust source regions; therefore, any increases in precipitation and rising concentrations of atmospheric CO$_2$ may contribute to decreased aridity and a possibly less dusty atmosphere in the future (Mahowald and Luo, 2003) (Fig. 35.5 path 7-8-9-11 inverse effect).

Increased precipitation over the oceans may increase the supply of a more “bioavailable,” dissolved aeolian Fe via wet deposition (Fig. 35.5 path 8-9-11 direct effect). During long range transport, dust particles undergo heterogeneous reactions at gas-solid-liquid interfaces involving pollution-derived substances that may lead to the increased solubility of aeolian Fe (Dentener et al., 1996; Meshidzke et al., 2003; Underwood et al., 2001). Photochemical reduction in more acidic cloud waters and precipitation promote dissolution of Fe in dust, leading to the production
of soluble Fe (II) (Siefert et al., 1999; Zhu et al., 1997; Zhuang et al., 1992). These natural processes potentially increase the amount of dissolved Fe in precipitation beyond the Fe solubility of a few percent directly measured from leaching of dry dust particles. Gao et al. (2003) suggest that the annual input of dissolved Fe by wet deposition accounts for 4–30% of the total aeolian Fe fluxes to the ocean. Hand et al., (Hand et al., 2004) reported that soluble iron associated with aerosols over the Atlantic and Pacific ranged from 0–45% in the PM$_{2.5}$ mode (particulate mass less than 2.5 µm diameter) and 0–87% in the coarse mode. These findings reveal that soluble iron could be a significant fraction of the total aeolian Fe entering the surface seawaters. Thus, increasing the supply of soluble aeolian Fe in the oceans through wet deposition may cause natural iron fertilization and induction of blooms by diazotrophic and other Fe-limited photosynthetic populations (Fig. 35.5 path 8–14). As such, variations in precipitation distributions, or changes in hydrological cycles caused by global warming, could alter carbon cycles in the ocean and contribute to the future climate change. The non-linear response of critical environmental factors over the land and oceans has restricted the characterization and predictive power of how precipitation and land-use patterns influence dust generation and subsequent aeolian Fe deposition to the oceans.

Changes in ocean stratification may further affect the degree of aeolian Fe demand by marine biota and the subsequent CO$_2$ fluxes (Le Quere et al., 2003). Intensified stratification in the upper ocean due to global warming could become more resistant to mixing by surface wind and prevent the flux of nutrients from deep waters to the surface layer (Sarmiento et al., 1998) (Fig. 35.5 path 13–14). This weakened mixing could also reduce the amount of recycled Fe brought from the deep ocean to the surface layer and, consequently, the supply of aeolian Fe to production in the surface ocean could be more critical in the future. This situation may occur both in the high-nutrient low-chlorophyll (HNLC) waters and also in the oligotrophic gyres, where there may be co-limitation of production by iron and phosphorous (Mills et al., 2004). In the eastern subtropical North Atlantic, where the impact of Saharan dust input is strong, utilization of nitrate supplied from dust deposition could be enhanced in the strongly stratified mixed layer during the late summer-fall period, thereby promoting phytoplankton growth (Neuer et al., 2004). Although primary productivity in the Sargasso Sea is often suggested to be phosphorus-limited, as the concentrations of dissolved inorganic phosphate in the surface waters are low (Wu et al., 2000), results from nutrient-addition bioassays in the tropical North Atlantic indicate that nitrogen fixation in this region is co-limited by iron and phosphorus (Mills et al., 2004). This finding suggests that even in a region where dust deposition is high and the amount of aeolian Fe should be sufficient, the rates of nitrogen fixation by high-Fe demanding organisms such as *Trichodesmium* spp. are still controlled by the supply of iron.

The combined effects of warming air temperatures and of enhanced stratification in the upper ocean may thus promote the dissolution of iron in dust and its subsequent bio-availability in the surface seawaters, possibly enhancing nitrogen fixation and export production. Future aeolian Fe input to the ocean may increase due to desertification enhanced by human activities or decrease by altered hydrological cycles (Fig. 35.5). These interactions and feedbacks could certainly play important roles in future nutrient and carbon cycles in the ocean.
3.2. Interactions of the oxygen and nitrogen cycles: Nitrification and denitrification

Changes in the ocean nitrogen budget linked to climate change have been discussed by several authors (Altabet et al., 1995; Broecker and Henderson, 1998; Falkowski, 1997; Ganeshram et al., 1995; Galbraith et al., Chapter 34 this volume). The coupling of the nitrogen and oxygen cycles via nitrification and denitrification has implications for the long-term chemical evolution of the ocean and atmosphere, the variations in the oceans' nitrogen inventory, and the nitrogen limitation of marine ecosystems over centennial and longer timescales. Yet, the interactions and feedbacks of the coupled nitrogen and oxygen cycles are not clearly understood. Consider a reduction of nitrogen fixation leading to a global decline in the fixed nitrogen budget of the ocean, increasing nitrogen limitation and reducing total export production. In response, deep ocean respiration decreases and the low oxygen zones shrink along with the integrated rate of denitrification. While there is the possibility for a stabilizing feedback (Codispoti, 1989; Deutsch et al., 2007), the processes of nitrogen fixation and denitrification are spatially (and temporally) disconnected; the former occurs predominantly in the surface, subtropical oceans while the latter occurs in the low oxygen zones of the water column and sediments (Fig. 35.1). Thus the rate of accumulation or the loss of nitrogen in the ocean must also be regulated by the processes of physical transport by which these regions are connected. In the modern ocean, the transport timescale between the subtropical Atlantic, a region of significant nitrogen fixation, and the low oxygen waters of the mid-depth Pacific basin, is on the order of hundreds of years leading to the potential for delayed responses in the feedback loop (e.g. Gruber, 2004; see Chapter 1, Gruber, this volume) and even to oscillatory behavior. A highly idealized box model of the coupled nitrogen and oxygen cycles (Fennel et al., 2005) suggests complex interactions (see following section) and illustrates how these interactions were likely critical in the development of global biogeochemical cycles on the early Earth. Factoring in possible links to the carbon pumps, climate and the global iron cycle suggests even richer, and more complex, possibilities for interactions of climate and biogeochemical cycles (e.g. Falkowski, 1997).

3.3. Ocean circulation and the global ocean cycles of nitrogen, oxygen and carbon

Ocean models forced by warmer atmospheric climates and coupled climate models in global warming scenarios suggest that weakened atmospheric temperature gradients and increased hydrological cycle might weaken the ventilation of the deep ocean (e.g. Zhang et al., 2001) and expand the regions of well-stratified, subtropical surface waters (e.g. Sarmiento et al., 2004). Consider the oceanic nitrogen cycle as represented simply in Fig. 35.6. Nitrogen is supplied to the surface ocean by ocean circulation, from depth, or by fixation in the surface. Depending upon the nature of the ecosystem and its efficiency at recycling, some fraction, \( N_{bio} \), of the deep ocean nitrogen, \( N_d \), is brought to depth through biological export of organic matter and remineralization. The remaining fraction, or the pre-formed nitrogen \( N_{pre} \), is brought to depth by physical advection or mixing from the surface layer. In regions
of low oxygen (including pore waters), denitrification and subsequent processes ultimately lead to a return to dinitrogen gas, and to nitrogen loss from the oceans.

The warming of the surface ocean will weaken the overturning circulation leading to shallower mixed-layer. Slower overturning suggests a longer residence time for waters in the surface, perhaps increasing the role of biological export relative to physical transport, favoring $N_{bio}$ over $N_{pre}$. In addition, a coincident warming of the surface waters and reduction of turbulent mixing in the surface mixed-layer might favor diazotrophs such as *Trichodesmium* (Capone, 2001; Karl *et al*., Chapter 16 this volume), enhance nitrogen fixation and work to increase the oceanic nitrogen budget, perhaps relieving nitrogen limitation. Consider, however, the coupled oxygen cycle. Shallower mixing and warming of the surface waters will reduce the physical supply of oxygen to the deep ocean and decrease the solubility of oxygen in the ventilated waters ($O_2^{pre}$ decreases) (which may further enhance nitrogenase activity and nitrogen-fixing species but will reduce productivity of other phytoplankton). Consequently, deep ocean oxygen concentrations, $O_2^{d}$, will likely decrease under such conditions, increasing the extent of regions of very low oxygen concentration in the water column (Zhang *et al*., 2001) and pore waters. In consequence, the global flux of denitrification is also likely to increase in response. The net effect of such a climate change on the oceanic nitrogen budget and productivity is not yet clear.

We examined the steady state response of the fully dynamic ocean nitrogen and oxygen cycles in our box model (see section Ib, Fennel *et al*., 2005) and an atmospheric mixing ratio of oxygen at 20% to prescribed variations in the ocean overturning, efficiency of nitrogen fixation and mean ocean phosphate loading. Starting from modern ocean conditions under which the oceanic N:P ratio is close to 16, varying the rate of overturning between 10 and 30 Sverdrups does little to the ocean nitrogen
inventory, or N:P, though the timescale for adjustment increases as overturning decreases. Changing the efficiency of nitrogen fixation, perhaps interpreted as reflecting changes in upper ocean stratification, has some interesting and counter-intuitive consequences; since the modern ocean has an N:P ratio close to 16, increasing the efficiency of nitrogen fixation does not significantly enhance the nitrogen budget, as the model assumes diazotrophs are not competitive unless nitrogen is limiting. However, for conditions where oceanic N:P is somewhat lower than 16, increasing nitrogen fixation can decrease the fixed nitrogen inventory of the deep ocean by enhancing export production, reducing deep ocean oxygen and increasing the rate of denitrification. Increasing the oceanic phosphorus inventory above today’s values also reduces N:P through a similar mechanism. These model explorations suggest there are critical non-linear interactions of between the C, N, O and P cycles which remain to be elucidated.

3.4. Implications for the global carbon cycle

What are the implications of such changes in the nitrogen cycle for the global carbon cycling and atmospheric pCO2? A biologically mediated net exchange of CO2 between the atmosphere and ocean can be brought about by changing the oceanic inventory of one or more limiting nutrients (Falkowski et al., 2003). If fixed inorganic nitrogen is limiting, then how sensitive is the atmospheric exchange of CO2 with the ocean to a change in the inventory of fixed inorganic nitrogen. To do this, we examined the effect of changing the nitrogen inventory (through biological fixation) in a coupled biogeochemical model. Following the treatment of the nitrogen and oxygen concentrations in the deep ocean, we may describe the ocean reservoir of dissolved inorganic carbon (DICd) as the sum of preformed (DICpre) and biological components (DICbio):

\[ \text{DIC}_d = \text{DIC}_{\text{pre}} + \text{DIC}_{\text{bio}} \] (35.4)

Here, the preformed pool includes contributions from the saturation concentration and the degree of disequilibrium at the point of subduction. The biological fraction has contributions from both soft-tissue and carbonate pumps (Volk and Hoffert, 1985). Following Brewer (Brewer, 1978) we may, as a first approximation, interpret the soft-tissue contribution to be related to the biological fraction of nitrogen by a fixed, Redfieldian stoichiometry:

\[ \text{DIC}_{\text{bio}} = R_{\text{CN}} N_{\text{bio}}. \]

If the ocean–atmosphere loading of carbon is fixed then, given these simplistic assumptions, enhancing the oceanic nitrogen budget and export production would lead to an enhancement of the oceans biological carbon pool at the expense of the atmosphere, reducing atmospheric pCO2 in proportion to the change in DIC according to the buffer factor, B:

\[ dpCO2/pCO2 \sim B \, dDIC/DIC \] (35.5)

(e.g. Bolin and Erickson, 1959) where \( B \sim O(10). \)
Accordingly, let us consider some climate change process which leads to an increase in fixed inorganic N by 1 μmol L\(^{-1}\) in the ocean interior, \(N_d\). Studies of the contemporary ocean suggest that the biological contribution \(N_{\text{bio}} \sim 0.3\ N_d\). Assuming this fraction stays constant and \(R_{CN} \sim 6\), this implies a corresponding 2 M increase in DIC\(_d\). The buffering relationship (2.5) suggests a modest associated decrease in atmospheric \(pCO_2\) (relative to 280 ppmv) of about 3 ppmv in the steady state (see Ito and Follows, 2004 for a more detailed analysis). If all of the extra nitrogen is in the biological fraction, the sensitivity of atmospheric \(CO_2\) to the mean ocean nitrogen concentration could be as much as 10 ppmv per micromole, relative to the pre-industrial state. It should be noted that this argument assumes no impact on the preformed component, DIC\(_{\text{pre}}\). It also assumes a fixed ocean-atmosphere carbon budget. However, modifications of the deep ocean carbonate system will ultimately lead to interactions with the sedimentary calcium carbonate reservoir, dampening the impact on atmospheric \(pCO_2\) (e.g. Bolin and Eriksson, 1959).

4. Summary and Conclusions

The history of Earth can be broadly divided into two “super eons.” The first 2.2 billion years were marked by biological innovation and experimentation, during which time all the major metabolic pathways evolved in prokaryotes. One of these pathways, oxygenic photosynthesis, would subsequently, over the second half of Earth’s history, give rise to aerobic metabolism, and ultimately permit the evolution of multicellular eukaryotes. The shift from mildly reducing to strongly oxidizing conditions was, beyond doubt, the most profound transition since the origin of life itself. Our analysis suggests that feedbacks between C, N, and O cycles helped prevent the oxidation of Earth in the Paleoproterozoic. This stabilizing feedback, which was ultimately over-ridden, led to the contemporary nitrogen cycle where nitrate, rather than ammonium, was the stable form of fixed inorganic nitrogen in the oceans. Barring some minor changes in the trace element composition in nitrogenases, the core proteins remained essentially unchanged following the transition to an oxidized atmosphere. In the contemporary ocean, approximately 20—30% of nitrogenase activity is inhibited at any moment in time by \(O_2\). This inhibition results in a negative feedback which constrains the upper level of \(O_2\) on Earth (Figs. 35.2 and 35.5).

Three central aspects of cyanobacterial nitrogen fixation remain curious. First, although some trace elements have been altered in the evolution of nitrogenases, the core proteins have remained virtually unchanged. In effect, nitrogenase, like RuBisCO, is a “frozen metabolic accident” (Shi et al., 2005). Either there is no substitute for the basic iron-sulfur cluster motif, or protein-protein interactions have severely constrained the evolutionary trajectory of this enzyme complex, or both. Whatever the reason, all nitrogenases are irreversibly inhibited by oxygen—and cyanobacteria are the only oxygenic photoautotrophs that contain the gene. Second, while there is abundant evidence of lateral transfer of nitrogenase genes between prokaryotes, in the endosymbiotic appropriation of cyanobacteria into heterotrophic hosts to photosynthetic eukaryotes, nitrogenases were lost. Indeed, there is no known eukaryote that contains an endogenous functional nitrogenase. It is not apparent whether this deficiency is
genetically insurmountable, but it is clear that (under present conditions) it made most aquatic eukaryotes dependent on cyanobacteria for fixed inorganic nitrogen. In oligotrophic subtropical and tropical oceans large chain-forming diatoms are found in association with the endosymbiotic heterocystous cyanobacterium, *Richelia intracellularis* that contribute significant amounts of fixed N to some systems (Capone et al., 1997; Carpenter et al., 1999; Foster and O’Mullan, Chapter 27, this volume; Janson et al., 1999; Venrick, 1974)(see Carpenter and Capone, Chapter 4, this volume). Yet, given the above dependency, the low diversity and limited biomass of endosymbiotic nitrogen fixing cyanobacteria harbored in eukaryotic host cells seems curious. Third, although free-living heterocystous cyanobacteria are abundant in lakes and brackish water ecosystems, they appear to be rare in the open ocean (Carpenter and Janson, 2001). Heterocysts are advantageous at lower temperature and salinities compared to *Trichodesmium* types, which dominate in the tropical oceans (Staal et al., 2003b). Yet, it is not clear why this, or some other biological innovation which facilitates nitrogen fixation under relatively high oxygen concentrations, is rare in temperate and polar oceanic ecosystems.

Clearly these issues will be the focus of research efforts over the next few decades because they lie at the heart of a fundamentally applied problem: why have we not been able to genetically engineer plants that feed and serve humans with the capability of fixing their own nitrogen? Should we overcome that major hurdle, we will have made a major transition in the impact of humans on the chemistry of Earth and its oceans.

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